Lactuca serriola L. (Asteraceae) **Prickly Lettuce**

Description. Biennial, sometimes persisting longer than 2 years, 5-20 dm tall, from a stout taproot. Stems 1 to several, erect, branches ascending, sharply hispid or prickly below, glabrous above. Leaves alternate, lower ones 5-20 cm long, oblong to oblance olate, deeply pinnately lobed, sagitatte, the lower lobes directed downward, prickly hispid, upper ones 2-15 cm long, clasping, ascending, somewhat glabrous or hispid along the lower margins. Heads in terminal panicles, ligulate (all flowers bilateral, corolla flat, 5-lobed), 8-12(16) mm long, cylindrical to conical, stalked. Phyllaries in 3-4 unequal series, oblong to lanceolate, glabrous, the outer much shorter than the inner. Flowers 10-20, corollas yellow. Achenes 3-3.5 mm long, ellipsoid to fusiform, ribbed, the beaks 3.5-4 mm long, the pappus soft-capillary, falling separately. (Barkley 1986, Clapham et al. 1962, Ferakova 1976, Ferris 1960, Gleason and Cronquist 1991, Stebbins 1993, Munz 1959).

Synonyms: The later name, *L. scariola* L. has been widely used for this species.

Note: A related species, L. saligna L. (willow lettuce), differs by having a narrow, spike-like inflorescence, subsessile heads, linear to lanceolate leaves with entire to lobed margins, but with the lobes restricted to near the base, and heads with 8-12 flowers. The closest relative of L. serriola appears to be L. sativa, the cultivated lettuce (de Vries 1996, 1997, Haqui and Godward 1984, Hill et al. 1996, Kesseli et al. 1991). Other species, including L. saligna, do not appear to be closely related and are completely cross-incompatible with either L. sativa and L. serriola (de Vries 1990, Kesseli et al. 1991). Lactuca serriola also shows a wide range of ecotypic variation, at least in Great Britain (Carter and Prince 1985, Prince and Carter 1985, Prince et al. 1985). At least 3 different races are reported from the North America (Barkley 1986, Gleason and Cronquist 1991).

Geographic distribution. Lactuca serriola is generally believed native to Europe, western Asia, and northern Africa (Clapham et al. 1962, Ferakova 1976, Munz 1959). It has become naturalized widely in North America, Australia, New Zealand, southern Africa, and South America (Arnold and De Wet 1993, Barkley 1986, Chapman 1991, Fernald 1950, Gleason and Cronquist 1991, Webb et al. 1988).

Lactuca serriola (as L. scariola) was first reported in the early 1890s from near Berkeley (Brandegee 1891) and northern California (Brandegee 1893). By the early 1900s, it had become more widely established throughout California (Robbins 1940). Lactuca serriola is known on all four northern Channel Islands (Junak et al. 1997) and it is reported from most counties in California (Anonymous 1998, Stebbins 1993).

Reproductive and vegetative biology. Lactuca serriola is self-compatible and experiences a high level of self-pollination (Stebbins 1958). Pollination in the related species, L. sativa, is effected primarily by small bees or hover flies (Syrphidae), at least in Britain (Watts 1958). Like other Asteraceae with a capillary pappus and small light seeds, Lactuca serriola has a relatively high level of dispersability (Anderson 1992, Sheldon and Burrows 1973). Dormancy in seeds of *L. serriola* is broken by a combination of light and moisture (Ellis et al 1989). Germination rates and dormancy also may be affected by

maturation date on long-lived plants and by patterns of variation in daily temperature (Gutterman 1992, 1994).

Ecological distribution. In both natural and naturalized geographic ranges, *Lactuca* serriola occurs on disturbed sites of waste areas, roadsides, open sites in grasslands, and abandoned fields (Amor 1986, Ferakova 1976, Gleason and Cronquist 1991, Munz 1959, Robbins et al. 1970, Webb et al. 1988).

Weed status. Lactuca serriola is not considered a serious noxious weed in agricultural or horticultural practice, at least at a global level (not listed by Holm et al. 1977), nor is it considered a noxious weed by the State Dept. of Food and Agriculture (Anonymous 1996). However, it is listed for the United States by Lorenzi and Jeffery (1987).

Microbial pathogens. Several viral, bacterial, and fungal pathogens have been reported from both L. serriola and L. sativa, including yellows virus, turnip mosaic potyvirus, Microdochium, Rhizomonas (corky root disease), Bremia (downy mildew), and Erysiphe (Galea and Price 1988, Lebeda 1984, 1986, 1989, 1994, McCreight 1987, O'Brien and Van Bruggen 1991, Van Bruggen et al. 1990). Natural variation in resistance to Rhizomonas and Bremia was reported by Bonnier et al. (1994), Brown and Michelmore (1988), Farrara et al. (1987), Maisonneuve (1994), Michelmore (1991), and Witsenboer et al. (1995) and to yellows virus (McCreight 1987). The primary purpose of these studies was to characterize genes conferring resistance, so that they could be used eventually in developing disease-resistant strains of cultivated lettuce.

Insect pathogens. White flies (*Bemisia* spp.) and noctuid moth larvae (*Trichoplusia ni*) are the only insects reported as causing damage to leaves of *Lactuca serriola* and *L*. sativa (Coudriet et al. 1986, Dussourd 1997, Summers et al. (1996) also showed that most whitefly larvae remained on the same plant on which they hatched.

Herbicide control. Lorenzi and Jeffery (1987) recommended the use of 2,4-D in waste places and along roadsides where other vegetation was desirable and the use of paraguat, bormacil, and sulfometuron for non-selective control. Resistance to several sulfonylurea herbicides (imidazolinone, imazapyr, imazaguin, imazethapyr) was reported by Mallory-Smith et al. (1990a, b) and Guttieri et al. (1992). Several references were found that report herbicide (primarily sulfonylurea) resistance, and its genetic basis, in wild strains of Lactuca serriola (e.g., Alcocer-Ruthling et al. 1992 a,b,c, Guttieri et al. 1992, Mallory-Smith et al. 1990a,b, 1993) The purpose of these studies was to search for genes conferring resistance, so that they could be used in developing herbicide-resistant strains of cultivated lettuce.

Literature Cited

Alcocer-Ruthling, M., D. Thill, and B. Shafi. 1992a. Differential competitiveness of sulfonylurea resistant and susceptible prickly lettuce (Lactuca serriola). Weed Technology. 6: 303-309.

- Alcocer-Ruthling, M., D. Thill, and C. Mallory-Smith. 1992b. Monitoring the occurrence of sulfonylurea-resistant prickly lettuce (Lactuca serriola). Weed Technology. 6: 437-440.
- Alcocer-Ruthling, M., D. Thill, and B. Shafi. 1992c. Seed biology of sulfonylurearesistant and -susceptible biotypes of prickly lettuce (*Lactuca serriola*). Weed Technology. 6: 858-864.
- Amor, R. 1986. Incidence and growth of prickly lettuce (*Lactuca serriola* L.) in dryland crops in the Victorian Wimmera. Plant Protection Quarterly. 1: 148-151.
- Anderson, M. 1992. An analysis of variability in seed settling velocities of several wind-dispersed Asteraceae. American Journal of Botany. 79: 1087-1091.
- Anonymous, 1996. Exotic pest plants of greatest ecological concern in California as of August 1996. California Exotic Pest Plant Council. 8 pp.
- Anonymous. 1998. California county flora database version 2, Santa Barbara Botanic Garden and USDA National Plants Data Center, Santa Barbara and New Orleans. URL = plants.usda.gov
- Arnold, T. and B. de Wet. 1993. Memoir 62. Plants of southern Africa: names and distribution. National Botanical Institute, Pretoria. 825 pp.
- Barkley, T. 1986. Asteraceae. pp. 838-1021. In Great Plains Flora Association. 1986. Flora of the Great Plains. University of Kansas, Lawrence. 1392 pp.
- Bonnier, F., K. Reinink, and R. Groenwald. 1994. Genetic analysis of *Lactuca* accessions with new major gene resistance to lettuce downy mildew. Phytopathology. 84: 462-468.
- Brandegee, K. 1891. Contribution to the knowledge of west american plants. I. Zoe 2: 75-83.
- Brandegee, T. 1893. New localities for California plants. Zoe 4: 148-159.
- Brown, P. and R. Michelmore. 1988. The genetics of corky root resistance in lettuce. Phytopathology. 78: 1145-1150.
- Carter, R. and S. Prince. 1985. The geographical distribution of prickly lettuce (*Lactuca* serriola). I. A general survey of its habitats and performance in Britain. Journal of Ecology. 73: 27-38.
- Chapman, A. 1991. Australian plan name index. K-P. Australian Government Publishing Service, Canberra. pp. 1711-2475.
- Clapham, A., T. Tutin, and E. Warburg. 1962. Flora of the British Isles. Cambridge University Press, Cambridge. 1269 pp.
- Coudriet, D. D. Meyerdirk, N. Prabhaker, and A. Kishaba. 1986. Bionomics of sweetpotato whitefly (Homoptera: Aleyrodidae) on weed hosts in the Imperial Valley, California. Environmental Entomology. 15: 1179-1183.
- de Vries, I. 1990. Crossing experiments of lettuce cultivars and species (*Lactuca* sect. Lactuca, Compositae). Plant Systematics and Evolution. 171: 233-248.
- de Vries, I. 1996. Characterization and identification of *Lactuca sativa* cultivars and wild relatives with SDS-electrophoresis (*Lactuca* sect. *Lactuca*, Compositae). Genetic Resources and Crop Evolution. 43: 193-202.
- de Vries, I.M. 1997. Origin and domestication of *Lactuca sativa* L. Genetic Resources and Crop Evolution. 44: 165-174.

- Dussourd, D. 1997. Plant exudates trigger leaf-trenching by cabbage loopers, Trichoplusia ni (Noctuidae). Oecologia. 112: 362-369.
- Ellis, R., T. Hong, and E. Roberts. 1989. Response of seed germination in three genera of Compositae to white light of varying photon flux density and photoperiod. Journal of Experimental Botany. 40: 13-22.
- Farrara, B., T. Ilott, and R. Michelmore. 1987. Genetic analysis of factors for resistance to downy mildew (Bremia lactucae) in species of lettuce (Lactuca sativa and L. serriola). Plant Pathology. 36: 499-514.
- Ferakova, V. 1976. Lactuca. pp. 328-331. In Tutin et al. (eds). Flora Europaea. Plantaginaceae to Compositae. Cambridge University Press, Cambridge. 505 pp.
- Fernald, M. 1950. Gray's Manual of Botany. Eighth Edition. American Book Company, New York. 1632 pp.
- Galea, V. and T. Price. 1988. Infection of lettuce by Microdochium panattonianum. Transactions of the British Mycological Society. 91: 419-425.
- Gleason, H. and A. Cronquist. 1991. Manual of the vascular plants of northeastern United States and Adjacent Canada. 2nd edition. New York Botanic Garden, Bronx. 910 pp.
- Gutterman, Y. 1992. Maturation dates affecting the germinability of *Lactuca serriola* L. achenes collected from a natural population in the Negev desert highlands. Germination under constant temperatures. Journal of Arid Environments. 22: 353-362.
- Gutterman, Y. 1994. Germinability under natural temperatures of *Lactuca serriola* L. achenes matured and collected on different dates from a natural population in the Negev Desert highlands. Journal of Arid Environments. 28: 117-127.
- Guttieri, M., C. Eberlein, C. Mallory-Smith, D. Thill, and D. Hoffman. 1992. DNA sequence variation in domain A of the acetolactate synthase genes of herbicideresistant and -susceptible weed biotypes. Weed Science. 40: 670-676.
- Haque, M. and M. Godward. 1985. Comparison between two genera, species and cultivars in Lactuceae. I. Karyotype analysis. Cytologia. 50: 725-738.
- Hill, M., H. Witsenboer, M. Zabeau, P. Vos, R. Kesseli, and R. Michelmore. 1996. PCR-based fingerprinting using AFLPs as a tool for studying genetiionships in spp. Theoretical and Applied Genetics. 93: 1202-1210.
- Holm, L., D. Plucknett, J. Pancho, and J. Herberger. 1977. The world's worst weeds: distribution and ecology. University Press of Hawaii, Honolulu. 609 pp.
- Junak, S., S. Chaney, R. Philbrick, and R. Clark. 1997. A checklist of vascular plants of Channel Islands National Park. Southwest Parks and Monuments Association, Tucson, AZ. 43 pp.
- Kesseli, R., O. Ochoa, and R. Michelmore. 1991. Variation at RFLP loci in *Lactuca* spp. and origin of cultivated lettuce (*L. sativa*). Genome. 34: 430-436.
- Lebeda, A. 1984. Response of differential cultivars of *Lactuca sativa* to *Bremia lactucae* isolates from Lactuca Serriola. Transactions of the British Mycological Society, 83: 491-494.
- Lebeda, A. 1986. Specificity of interactions between wild *Lactuca* spp. and *Bremia* lactucae isolates from Lactuca serriola. Phytopathologische Zeitschrift. 117: 54-64.
- Lebeda, A. 1989. Response of lettuce cultivars carrying the resistance gene

- Dm11 to isolates of Bremia lactucae from Lactuca serriola. Zeitschrift fur Pflanzenzuchtung. 102: 311-316.
- Lebeda, A. 1990. The location of sources of field resistance to *Bremia lactucae* in wild Lactuca species. Zeitschrift für Pflanzenzuchtung. 105: 75-77.
- Lebeda, A. 1994. Evaluation of wild *Lactuca* species for resistance of natural infection of powdery mildew (*Ervsiphe cichoracearum*). Genetic resources and crop evolution. 41: 55-57.
- Lorenzi, H. and L. Jeffery. 1987. Weeds of the United States and their control. Van Nostrand Company, New York. 355 pp.
- Mallory-Smith, C., D. Thill, and M. Dial. 1990a. Identification of sulfonylurea herbicideresistant prickly lettuce (Lactuca serriola). Weed Technology. 4: 163-168.
- Mallory-Smith, C., D. Thill, M. Dial, and R. Zemetra. 1990b. Inheritance of sulfonylurea herbicide resistance in Lactuca spp. Weed Technology 4: 787-790.
- Mallory-Smith, C., D. Thill, and M. Dial. 1993. ID-BR1: sulfonylurea herbicide-resistant lettuce germplasm. HortScience. 28: 63-64.
- Maisonneuve, B., Y. Bellec, P. Anderson, and R. Michelmore. 1994. Rapid mapping of two genes for resistancea to downy mildew from *Lactuca serriola* to existing clusters of resistance genes. Theoretical and Applied Genetics. 89: 96-104.
- McCreight, J. 1987. Resistance in wild lettuce to lettuce infectious yellows virus. HortScience, 22: 640-642.
- Michelmore, R., I. Paran, and R. Kesseli. 1991. Identification of markers linked to disease-resistance genes by bulked segregant analysis: a rapid method to detect markers in specific genomic regions by using segregating populations. Proceedings of the National Academy of Sciences. 88: 9828-9832.
- Munz, P. 1959. A flora of California. University of California Press, Berkeley. 1681 pp.
- O'Brien, R. and A. Van Bruggen. 1991. Populations of Rhizomonas suberifaciens on roots of host and nonhost plants. Phytopathology. 81: 1034-1038.
- Prince, S. and R. Carter. 1985. The geographical distribution of prickly lettuce (*Lactuca* serriola). III. Its performance in transplant sites beyond its distribution limit in Britain. Journal of Ecology. 73: 49-64.
- Prince, S., R. Carter, and K. Dancy. 1985. The geographical distribution of prickly lettuce (Lactuca serriola). II. Characteristics of populations near its distribution limit in Britain. Journal of Ecology. 73: 39-48.
- Richards, A. 1978. The pollination of flowers by insects. Linnean Society Symposium Series 6: 1-213. Academic Press, London.
- Robbins, W. 1940. Alien plants growing without cultivation in California. Agricultural Experiment Station. Bulletin 637. University of California, Berkeley. 128 pp.
- Robbins, W., M. Bellue, and W. Ball. 1970. Weeds of California. Documents and Publications, Sacramento, California, 547 pp.
- Sheldon, J. and F. Burrows. 1973. The dispersal effectiveness of the achene-pappus units of selected Compositae in steady winds with convection. New Phytologist 72: 665-675.
- Stebbins, G. 1993. *Lactuca*. p. 296. In Hickman, J. (ed.) The Jepson Manual: higher plants of California. University of California Press, Berkeley. 1400 pp.

- Summers, C., A. Newton, and D. Estrada. 1996. Intraplant and interplant movement of Bemisia argentifolii (Homoptera: Aleyrodidae) crawlers. Environmental entomology. 25: 1360-1364.
- Van Bruggen, A., P. Brown, and K. Jochimsen. 1990. Host range of *Rhizomonas* suberifaciens, the causal agent of corky root of lettuce. Plant Disease. 74: 581-584.
- Webb, C., W. Sykes, and P. Garnock-Jones. 1988. Flora of New Zealand. Volume 4. Naturalized pteridophytes, gymnosperms, dicotyledons. Department of Scientific and Industrial Research, Christchurch. 1365 pp.
- Witsenboer, H., R. Kesseli, M. Fortin, M. Stranghellini, and R. Michelmore. 1995. Sources and genetic structure of a cluster of genes for resistance to three pathogens in lettuce. Theoretical and Applied Genetics. 91: 178-188.